

Evaluation of aquatic herbicides applied under simulated drawdown conditions for control of dioecious hydrilla subterranean turions

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ABSTRACT

The use of drawdowns for aquatic weed control has been practiced for many years across the United States. However, highly persistent aquatic weeds such as hydrilla are not thoroughly controlled from drawdown conditions because of the production of desiccation-tolerant propagules like turions. Therefore, an experiment was conducted in Florida to evaluate hydrilla suppression following herbicide treatments during simulated drawdown conditions. Herbicide treatments included the 0.5, 1, and 2× foliar or drawdown rates of bispyribac-sodium, imazamox, penoxsulam, flumioxazin, topramezone, and florypyrauxifen-benzyl. Treatments were made to dishpans planted with hydrilla subterranean turions and transferred to a pond 7 d after treatment (DAT). Bispyribac-sodium and imazamox provided 95 to 100% control in both experimental runs across all rates. Penoxsulam, topramezone, and florypyrauxifen-benzyl did not perform consistently in both experimental runs; however, all provided excellent hydrilla biomass suppression in one of two site-years. Flumioxazin performed consistently across both experimental runs resulting in 70 to 89% biomass reduction. These results indicate high potential utility of currently registered aquatic herbicides for hydrilla control when applied under drawdown conditions.

Key words: chemical control, dewatered, growth retardant

INTRODUCTION

Substantial seasonal variations in water levels can drive shifts in aquatic plant community abundance and composition, population dynamics of sport fish, and nutrient levels (Chow-Fraser et al. 1998, Havens et al. 2004, 2005, Van Geest et al. 2005, Sharip et al. 2012). However, many systems in the United States today are stabilized by water control structures for flood control and water supply (Arnold 1988). The use of drawdowns is a widely accepted tool for both fisheries and aquatic plant management and has been practiced since the 1940s in systems where it is

feasible (Hinman and Hess 1949, Heman et al. 1969, Richardson 1975, Cooke 1980, Nagid et al. 2015). Drawdowns in the United States are most common during fall and winter months (Cooke 1980). Aquatic plant control from drawdowns is achieved primarily through desiccation in southern states; whereas in the northern United States, greater aquatic plant control from drawdowns has been attributed to the combined effects of desiccation and freezing temperatures (Richardson 1975, Reimer 1984, Wersal et al. 2013, Bellaud 2021), though insulation by snow cover in northern states may provide propagule protection from freezing temperatures.

The efficacy of drawdowns on aquatic plants is species-specific (Cooke 1980). Submersed plants that primarily reproduce from root crowns and vegetative fragments (e.g., *Myriophyllum* spp., *Cabomba* spp., *Ceratophyllum* spp., and *Egeria* spp.) are typically more susceptible to drawdowns (Cooke 1980, Bellaud 2021). For example, egeria (*Egeria densa* Planch.) and cabomba (*Cabomba caroliniana* A. Gray var *pulcherrima* Harper) were effectively controlled for a full growing season following a 4-mo drawdown during fall and winter months in Louisiana (Manning and Johnson 1975). In addition, greater weed control following drawdowns has been suggested to occur when higher turbidity, water levels, and lower temperatures occur during spring refill periods (Manning and Sanders 1975). Species that produce abundant seeds, turions, or winter buds such as hydrilla [*Hydrilla verticillata* (L.f.) Royle], naiads (*Najas* spp.), and pondweeds (*Potamogeton* spp.) are generally not controlled by drawdowns and often increase in coverage and density following refill due to reduced competition from slower colonizing species (Kadlec 1962, Nichols 1975, Cooke 1980, Poovey and Kay 1998). Two growing seasons after a 3-mo drawdown at Lake Ocklawaha, Florida, hydrilla had increased from 10 ha (pre-drawdown) to more than 650 ha (Hestand and Carter 1974, 1975). Drawdown effects are very brief for hydrilla in particular because of its persistent subterranean turions that commonly found at densities as high as 1,000 turions m⁻² (Haller and Sutton 1975, Harlan et al. 1985, Sutton and Portier 1985, Van and Steward 1990). Following a 12-mo drawdown of Lake Ray Roberts in Texas, no reductions in hydrilla subterranean turions or viability were observed (Doyle and Smart 2001). Furthermore, hydrilla subterranean turions consist of 50 to 60% water by weight and will maintain viability unless water content is reduced to 10% (Doyle and Smart 2001).

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A potential management technique to improve the efficacy of drawdowns on troublesome submersed aquatic species such as hydrilla is to integrate herbicide application during dewatered periods. This technique is practiced in some areas of the United States but is not well represented in the scientific literature. Manning and Johnson (1975) reported hydrilla recruitment from turions following dewatered treatments of 2,4-D amine at 6 kg ha⁻¹. Steward (1980) evaluated several herbicides and growth regulators on hydrilla subterranean turions and discovered that only fenac (2,3,6-trichlorophenyl acetic acid) and dichlobenil (2,6-dichlorobenzonitrile) suppressed hydrilla regrowth from these structures in laboratory and pond studies. Fenac, dichlobenil, and silvex [2-(2,4,5-trichlorophenoxy)-propionic acid] were also reported to control *Potamogeton* species in western irrigation canals (Frank et al. 1963). However, fenac, dichlobenil, and silvex were not reregistered for aquatic use in the late 1970s and early 1980s. Published studies on drawdown applications of currently registered aquatic herbicides is limited to sago pondweed (*Stuckenia pectinata*) and flowering rush (*Butomus umbellatus*). Vassios (2010) reported 74 to 77% biomass reduction 30 d after treatment of sago pondweed tubers in drawdown applications of imazamox, imazapyr, penoxsulam, fluridone, or flumioxazin. Likewise, drawdown applications of imazapyr and imazamox reduced flowering rush in Idaho (Getsinger and Madsen 2021). Since drawdowns alone provide only short-term control of hydrilla, and no published data exist in the literature on the efficacy of currently registered aquatic herbicides, the objective of this work was to evaluate currently registered aquatic herbicides for hydrilla control when applied under drawdown conditions. If hydrilla growth can be retarded and the longevity of control from a drawdown can be extended by a dewatered herbicide application, this practice could become a practical management option where possible.

MATERIALS AND METHODS

An experiment was conducted from winter to spring of 2020 (initiated in January) and repeated in 2022 (initiated in February) to evaluate the use of aquatic herbicides applied in a simulated drawdown to determine their impacts on hydrilla growth from subterranean turions. The experiment was set up as a completely randomized design with four replications and a factorial arrangement of treatments plus a nontreated control. Factors consisted of herbicide (bispiribac-sodium¹, imazamox², penoxsulam³, flumioxazin⁴, topamezone⁵, and florypyrauxifen-benzyl⁶) and rate (0.5×, 1×, or 2× the maximum foliar or dewatered use rate) (Table 1). In the second experimental run, three additional herbicide active ingredients were included and replicated at single (1×) use rates: potassium salt of endothall⁷, dichlobenil⁸, and imazapyr⁹. Potassium salt of endothall was included because, to our knowledge, this formulation had never been evaluated under drawdown conditions. A use rate of 7.1 kg ai ha⁻¹ was used based on a previous Florida 24C label for foliar applications to control floating plants (G. Adrian, pers. comm.). Imazapyr was included because it has a 24C label for drawdown use in Florida as well as

TABLE 1. HYDRILLA BIOMASS REDUCTION FOLLOWING SIMULATED DRAWDOWN HERBICIDE TREATMENTS.

Herbicide	Rate (g ai ha ⁻¹)	% Biomass reduction ^{1,2}	
		2020	2022
Bispiribac-sodium	56	99 a	95 ab
	112	100 a	95 ab
	224	100 a	95 ab
Imazamox	561	96 ab	96 ab
	1,123	100 a	96 ab
	2,245	99 a	96 ab
Penoxsulam	98	94 ab	41 f
	197	95 ab	43 f
	393	99 a	44 f
Flumioxazin	211	71 c	74 de
	422	70 c	79 cd
	844	81 bc	89 abc
Topamezone	195	7 f*	61 e
	390	10 f*	76 cd
	779	31 e	94 ab
Florypyrauxifen-benzyl	29	7 f*	93 ab
	58	53 d	94 ab
	117	87 ab	100 a

¹Means followed by the same letter are not different according to Fisher's LSD test ($\alpha = 0.05$).

²Asterisk signifies mean is not significantly different from the nontreated control (biomass of nontreated controls was 376 g dishpan⁻¹ in 2021 and 431 g dishpan⁻¹ in 2022).

recently reported efficacy on flowering rush (*Butomus umbellatus* L.) when applied during drawdown conditions in Montana (Getsinger and Madsen 2021). Lastly, dichlobenil was included as a historical reference treatment based on its previous aquatic registration and high levels of efficacy on hydrilla (Taylor 1966, 1967, Steward 1980).

Plastic dishpans¹⁰ (11.4 L; 34 cm long, 29 cm wide, and 14 cm deep) were filled with 10.5 cm of masonry sand (no organic matter; pH 7.5 to 8.5) amended with slow-release fertilizer (8-6-12, Osmocote Classic)¹¹ at a rate of 1.5 g kg⁻¹ of dry sand. Masonry sand was used to provide the best-case scenario for herbicide availability to subterranean turions by minimizing soil sorption. Drain holes (7 mm) were drilled in each bottom corner of the dish pans for excess water to drain during incorporation and to fill from the bottom with water in the grow out pond during refill. Dishpans were then placed on ground cloth in an unheated greenhouse in Gainesville, FL. Fluridone-susceptible dioecious hydrilla subterranean turions were collected and screened for maturity from the sediments of a shallow pond at the University of Florida (29°43'17.55"N; 82°25'2.28"W). After quiescent subterranean turions were collected, they were placed in a sealed plastic bag to minimize exposure to atmospheric oxygen and kept in the dark at room temperature overnight prior to planting. The purpose of this special handling was to keep the subterranean turions from breaking dormancy and sprouting before being placed in the experimental grow out ponds (Netherland 1997).

Approximately 18 h after subterranean turion collection, dish pans were watered with 13 mm of simulated rainfall, and 20 subterranean turions per dishpan were evenly placed on the surface of the sand, then covered with an additional 2.5 cm of moist sand. Dishpans were then treated with the herbicide treatments and randomly placed back in

the greenhouse. In 2022, three additional herbicides at a single maximum rate were included for reference. Herbicides were applied with a handheld boom equipped with XR11005 nozzles¹² and a CO₂-pressurized backpack sprayer calibrated to deliver 934 L ha⁻¹ at 276 kPa. Following application, 13 mm of overhead irrigation was applied to all dishpans. Dishpans were left in the greenhouse and received irrigation as needed to keep subterranean turions moist until placement in South Florida Water Management District research ponds (0.3 ha; average water pH ~7) near Wellington, FL, 7 d after treatment (26°37'27.81"N; 80°25'33.06"W) (Guardo et al. 1995). In 2020 dishpans received an additional 13 mm of irrigation at 3 d after treatment (DAT); however, in 2022 dishpans were irrigated with 13 mm at 3 and 6 DAT. Average daily temperatures during greenhouse periods were 9.6 C (SD = 3.2) and 18.0 C (SD = 3.4) in 2020 and 2022, respectively.

Seven days after herbicide application, dishpans were randomly placed on ground cloth approximately 40 to 50 cm apart on the bottom of the dry pond. In-flow water pumps were then turned on (18.2 L s⁻¹) to slowly fill the pond (dishpans were submersed in approximately 3 h) and provide a continuous hydraulic retention time of approximately 48 h. The pond was maintained at a water depth of 1.4 m with continuous water flow. After 18 (run 1) and 16 (run 2) wk, all dishpans were collected and hydrilla biomass harvested, rinsed, placed in mesh laundry bags, and dried in a forced air dryer at 60 C for 5 d and weighed (Figure 1).

A separate reference sprouting experiment was set up to determine the percentage viability of collected subterranean turions used in pond experiment. In runs 1 and 2, 160 and 40 mature subterranean turions, respectively, were randomly collected from batches used to plant dishpans. Subterranean turions were then placed in 500 ml beakers (20 turions per beaker) containing 350 ml of water. Beakers were placed on a laboratory bench at room temperature and ambient fluorescent lighting and monitored for 4 wk. Sprouted tubers were counted and removed and water was changed one to two times per week.

Hydrilla biomass data were normalized to the nontreated control and arcsine square-root transformed to improve heterogeneity of variance; however, back-transformed means are presented for clarity. Data were subject to ANOVA to test for main effects and interactions in R (version 3.6.1; R Core Team 2019). Where significant effects were detected, means were separated using Fisher's LSD test (0.05) under the "agricolae" package in R (Mendiburu 2019).

RESULTS AND DISCUSSION

Cumulative subterranean turion sprouting rates in the reference sprouting experiment were ≥80% over the 4-wk period. Additionally, dry aboveground biomass from nontreated controls in the field experiment resulted in 376 and 431 g dishpan⁻¹ in 2020 and 2022, respectively, which indicated excellent growing conditions during the experiment (Table 1). A significant interaction between main effects and experimental run were detected; therefore, experimental runs were analyzed separately (Table 1). Likewise, there



Figure 1. Run 1 (2020) dishpans 7 wk after treatment.

was a significant interaction between main effects (herbicide and rate) in both years.

Bispyribac-sodium and imazamox performed similarly and reduced hydrilla biomass 95 to 100% across all rates and in both experimental runs (Table 1). These two herbicides were the only products tested that provided >95% biomass reduction consistently across both years. The primary mechanism for both bispyribac-sodium and imazamox degradation in soil and water is microbial degradation (Aichele and Penner 2005, Kalsi and Kaur 2019). Half-lives ranging from 4 to 41 d have been reported for bispyribac-sodium in terrestrial soils depending on soil pH, organic matter, temperature, and moisture (Chirukuri and Atmakuru 2015, Kalsi and Kaur 2019). Likewise, imazamox has been shown to have a similar soil persistence to bispyribac-sodium in terrestrial soils, especially under low moisture and low pH conditions (Cobucci et al. 1998, Bresnahan et al. 2002, Aichele and Penner 2005). These characteristics in combination with both herbicides' ability to be absorbed by belowground plant tissues have been documented in terrestrial systems to provide varying levels of pre-emergence weed control and carryover injury to subsequent crops (Cobucci et al. 1998, Ball et al. 2003, Pannacci et al. 2006, Teuton et al. 2007, Rood et al. 2012, Lycan and Hart 2017a, b, Harmanci et al. 2024). However, in the current study soil moisture was not limiting, pH was likely between 7.5 and 8.5, and microbial activity prior to placement in test ponds was likely low. Therefore, imazamox and bispyribac were most likely highly available in the soil with minimal microbial degradation.

Penoxsulam performed very well in 2020 resulting in 94 to 99% biomass reduction across all rates tested (Table 1). However, in 2022 hydrilla biomass reduction did not exceed 44% from penoxsulam applied at the 2× rate. Though penoxsulam's terrestrial use patterns are largely for post-emergent weed control in rice systems, residual weed control of up to 3 wk after treatment has been reported (Mann 2015, Turra et al. 2023). In the current study, inconsistent results on hydrilla suppression may be due to slightly lower

air temperatures in 2020 (9.6 C) than in 2022 (18.0 C) during the period subterranean turions were exposed to herbicide prior to inundation. Penoxsulam has been shown to have a highly variable half-life in terrestrial soils with reported values ranging from 2 to 30 d depending on soil type (Jabusch and Tjeerdema 2006, Monika et al. 2017, Kaur and Kaur 2018). Kaur and Kaur (2018) reported increased penoxsulam dissipation in terrestrial soils when soil moisture and temperature increased. Thus, warmer pre-inundation temperatures in 2022 coupled with the use of coarse masonry sand may have contributed to faster penoxsulam degradation resulting in shorter exposure times to subterranean turions. Furthermore, penoxsulam degradation is faster under more alkaline conditions, which was likely the case in the current study which utilized sand with pH ranging from 7.5 to 8.5 (Joshi and Srivastava 2019).

Hydrilla biomass reduction from drawdown applications of flumioxazin was relatively consistent between years ranging from 70 to 81% reduction in 2020 and 74 to 89% reduction in 2022 (Table 1). Flumioxazin, which is primarily degraded through pH-mediated hydrolysis, can vary greatly in terms soil persistence with terrestrial soil half-lives reported to 10 to 32 d (Ferrell and Vencill 2003, Alister et al. 2008, Mueller et al. 2014, Eason et al. 2022). However, the consistency of flumioxazin across both experimental runs in the current experiment suggests that pH conditions were similar (soil = 7.5 to 8.5; water = 7) in both years and degradation may have not been affected by temperature. Additionally, flumioxazin's efficacy on hydrilla subterranean turions is reasonable considering its extensive pre-emergent use patterns in terrestrial systems (Askew et al. 2002, Grey et al. 2003, Grichar 2006, Richardson and Zandstra 2006, 2009, Mahoney et al. 2014, Herrmann et al. 2017).

Topramezone did not reduce hydrilla biomass compared to the nontreated control when applied at the 0.5× and 1× rates in 2020 (Table 1). However, a topramezone rate response was observed in 2022, and hydrilla biomass was reduced 61 to 94%. In terrestrial systems, topramezone is primarily utilized for post-emergence weed control but does possess some soil residual activity on sensitive species (Rahman et al. 2015). In terrestrial soils, topramezone bioavailability is lower in sandy soils with lower cation exchange capacities and at higher pH (Ramanathan et al. 2023). Considering the use of masonry sand in the current experiment, soil interactions with topramezone should have been identical in both experimental runs with the exception of an additional irrigation event in 2022. However, topramezone's mechanism of action (inhibition of 4-hydroxyphenylpyruvate dioxygenase) requires treated sensitive plants to be exposed to light for bleaching symptoms to appear (Grossman and Ehrhardt 2007). Thus, delayed hydrilla subterranean turion emergence coupled with minimal root formation for herbicide uptake in 2020 due to cooler temperatures (9.6 C) may be an explanation for the observed inconsistent results.

Hydrilla biomass reductions from drawdown applications of florypyrauxifen-benzyl were inconsistent among experimental runs (Table 1). In 2020 a 2× rate was required to achieve 87% biomass reduction; however, in 2022 biomass reduction was at least 93% at the 0.5× rate. Despite the observed inconsistencies, results from 2022 are encouraging

TABLE 2. HYDRILLA BIOMASS REDUCTION FOLLOWING SIMULATED DRAWDOWN HERBICIDE TREATMENTS IN 2022.

Herbicide	Rate (g ai ha ⁻¹)	% Biomass reduction ^{1,2,3} (2022)
Potassium salt of endothall	7,100	44 c
Dichlobenil	11,200	100 a
Imazapyr	1,682	93 ab

¹Biomass of nontreated control was 431 g dishpan⁻¹.

²Means followed by the same letter are not different according to Fisher's LSD test ($\alpha = 0.05$).

³Asterisk signifies mean is not significantly different from the nontreated control.

considering that florypyrauxifen-benzyl is primarily utilized as a post-emergent option with minimal pre-emergence activity (Miller and Norsworthy 2018, Wright et al. 2020). A potential explanation for inconsistencies among years may again be due to warmer greenhouse temperatures in 2022 that likely promoted earlier turion sprouting and emergence resulting in a slight increase in time for herbicide uptake and activity prior to inundation in test ponds.

Reductions in hydrilla biomass in 2022 from drawdown applications of potassium salt of endothall, dichlobenil, and imazapyr were 44, 100, and 93%, respectively (Table 2). Despite endothall having previous soil-applied terrestrial use patterns (Turgeon et al. 1972a,b), application to aquatic sediments with significant microbial activity for degradation likely limits its effectiveness (Simsman and Chester 1975, Islam et al. 2018). Likewise, Steward (1969) reported minimal phytotoxicity of the cocoamine salt of endothall on hydrilla turions. However, sand typically has very low microbial activity, which suggests that the lack of hydrilla suppression observed from endothall was not due to limited availability. Dichlobenil resulted in 100% reduction of hydrilla biomass, which speaks to its previous aquatic registration and dewatered use patterns (Table 2; Hiltbran 1966). Dichlobenil was first registered more than 60 yr ago for aquatic use and was not reregistered due to its persistence and very high use rates. Its inclusion in the current study suggests that the methodology is suitable since results were similar to those of Steward (1980) (Montgomery et al. 1972, Clausen et al. 2004). Isoxaben and indaziflam, two newer herbicides that share the same mode of action as dichlobenil (inhibition of cellulose synthesis), may be candidates for future aquatic drawdown evaluation. Despite not being labeled for in-water applications or having significant activity on hydrilla, drawdown application of imazapyr resulted in 93% biomass reduction (Table 2; Nelson et al. 1991, Patten 2003). Among aquatic weed scientists, imazapyr is largely considered inactive on submersed aquatic plants due to its relatively short aqueous half-life (Bajrai et al. 2017, Schreiber et al. 2017, Dugdale et al. 2020). However, the hydrilla subterranean turion response observed in the current experiment is more in accordance with reports of its substantial pre-emergent activity in terrestrial systems (Bovey and Senseman 1998, Harrington et al. 1998, Ulbrich et al. 2005, Douglass et al. 2016, Zaccaro et al. 2018).

Overall, hydrilla biomass reduction levels were greatest and most consistent across years following drawdown treatments of bispyribac-sodium and imazamox. These two herbicides also reduced hydrilla biomass similarly at 0.5× and

1× rates, which gives extraordinary forgiveness to applicators for error that cannot be controlled under field conditions. Additionally, bispyribac-sodium and imazamox in both experimental runs, and florypyrauxifen-benzyl in 2022 provided hydrilla control like that of the historical standard active ingredient, dichlobenil.

One of the most compelling aspects of hydrilla subterranean turion response to herbicides under drawdown conditions is how different it can be from whole-plant response from in-water exposure. For example, the slow-acting herbicides tested require several months of exposure time to provide hydrilla control from in-water application (Shearer and Nelson 2009, Getsinger et al. 2011, Sartain 2014, Wood and Netherland 2019). However, in the current experiment, these slow-acting active ingredients provided exceptional hydrilla control after exposing subterranean turions for only 7 d, although theoretical concentrations in the substrate were much higher than typical in-water use rates. Furthermore, this is only under the assumption that once dishpans were covered with water in ponds 7 d after treatment, herbicide concentrations in soil pore water were rapidly reduced due to the diffusive pressure of the surrounding water column.

Much further research is needed on the utility and operational refinement of drawdown herbicide applications. Additional rate-response experiments are recommended to determine optimal and cost-effective doses to extend hydrilla suppression. Likewise, further investigations of the drivers of herbicide consistency and associated environmental conditions are needed as drawdown herbicide applications can be highly variable (Madsen et al. 2017). Similarly, little to no published data exist on drawdown herbicide application selectivity toward key desirable plant species. Future work will also investigate drawdown herbicide efficacy on hydrilla under operational conditions.

In summary, Steward (1980) reported that underground structures of hydrilla such as subterranean turions, rhizomes, and root crowns are responsible for rapid reinfestation following control measures. Therefore, increased effectiveness of hydrilla control practices is dependent on the ability to reduced regrowth potential of underground structures. Based on the observations in the current experiment, there are several currently registered aquatic herbicides with great potential for hydrilla control from drawdown applications.

SOURCES OF MATERIALS

- ¹Tradewind®, Valent U.S.A. Corporation, Walnut Creek, CA 94596.
- ²Clearcast®, SePRO Corporation, Carmel, IN 46032.
- ³Galleon® SC, SePRO Corporation, Carmel, IN 46032.
- ⁴Clipper® SC, Nufarm Inc., Alsip, IL 60803.
- ⁵Oasis®, SePRO Corporation, Carmel, IN 46032.
- ⁶Procellacor® SC, SePRO Corporation, Carmel, IN 46032.
- ⁷Aquathol® K, UPL NA INC, King of Prussia, PA 19406.
- ⁸Casoron® 4G, UPL NA INC, King of Prussia, PA 19406.
- ⁹Habitat®, SePRO Corporation, Carmel, IN 46032.
- ¹⁰Rubbermaid® dishpan, Newell Brands, Atlanta, GA 30328.
- ¹¹Osmocote Smart Release 15-9-12®, Scotts, Marysville, OH 43041.
- ¹²XR 11005 nozzles, TeeJet® Technologies, Springfield, IL 62703.

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LITERATURE CITED

- Aichele TM, Penner D. 2005. Adsorption, desorption, and degradation of imidazolinones in soil. *Weed Technol.* 19(1):154–159.
- Alister C, Rojas S, Gomez P, Kogan M. 2008. Dissipation and movement of flumioxazin in soil at four field sites in Chile. *Pest Manag. Sci.* 64(5):579–583.
- Arnold JL. 1988. The evolution of the 1936 flood control act. Office of History, U.S. Army Corps of Engineers. 123 pp.
- Askew SD, Wilcut JW, Cranmer JR. 2002. Cotton (*Gossypium hirsutum*) and weed response to flumioxazin applied preplant and postemergence directed. *Weed Technol.* 16(1):184–190.
- Bajrai FSM, Ismail BS, Mardiana-Jansar K, Omar R. 2017. Persistence of imazapic and imazapyr in paddy soil and water. *Int. J. Adv. Agric. Environ. Eng.* 4(1):12:15.
- Ball DA, Yenish JP, Alby T. 2003. Effect of imazamox soil persistence on dryland rotational crops. *Weed Technol.* 17(1):161–165.
- Bellaud MD. 2021. Cultural and physical control of aquatic weeds, pp. 129–132. In: L. A. Gettys, W. T. Haller and D. G. Petty (eds.). *Biology and control of aquatic plants: A best management practices handbook*. 4th ed. Aquatic Ecosystem Restoration Foundation, Marietta, GA. 224 pp.
- Bovey RW, Senseman SA. 1998. Response of food and forage crops to soil-applied imazapyr. *Weed Sci.* 46(5):614–617.
- Bresnahan G, Dexter A, Koskinen W, Lueschen W. 2002. Influence of soil pH-sorption interactions on the carry-over of fresh and aged soil residues of imazamox. *Weed Res.* 42(1):45–51.
- Chirukuri R, Atmakuru R. 2015. Sorption characteristics and persistence of herbicide bispyribac sodium in different global soils. *Chemosphere* 138:932–939.
- Chow-Fraser P, Loughheed V, Le Thiec V, Crosbie B, Simser L, Lord J. 1998. Long-term response of the biotic community to fluctuating water levels and changes in water quality in Cootes Paradise Marsh, a degraded coastal wetland of Lake Ontario. *Wetl. Ecol. Manag.* 6:19–42.
- Clausen L, Larsen F, Albrechtsen HJ. 2004. Sorption of the herbicide dichlobenil and the metabolite 2,6-dichlorobenzamide on soils and aquifer sediments. *Environ. Sci. Technol.* 38(17):4510–4518.
- Cobucci T, Prates HT, Falcao CLM, Rezende MMV. 1998. Effect of imazamox, fomesafen, and acifluorfen soil residue on rotational crops. *Weed Sci.* 46(2):258–263.
- Cooke GD. 1980. Lake level drawdown as a macrophyte control technique. *Water Resour. Bull.* 16:317–322.
- Douglass CH, Nissen SJ, Meiman PJ, Kniss AR. 2016. Impacts of imazapyr and triclopyr soil residues on the growth of several restoration species. *Rangeland Ecol. Manag.* 69(3):199–205.
- Doyle RR, Smart RM. 2001. Effects of drawdowns and desiccation on subterranean turions of hydrilla, an exotic aquatic weed. *Weed Sci.* 49:135–140.
- Dugdale TM, Butler KL, Finlay MJ, Liu Z, Rees DB, Clements D. 2020. Residues and dissipation of the herbicide imazapyr after operational use in irrigation water. *Int. J. Environ. Res. Public Health* 17(7):2421.
- Eason K, Grey T, Cabrera M, Basinger N, Hurdle N. 2022. Assessment of flumioxazin soil behavior and thermal stability in aqueous solutions. *Chemosphere* 288(2):132477.
- Ferrell JA, Vencill WK. 2003. Flumioxazin soil persistence and mineralization in laboratory experiments. *J. Agric. Food Chem.* 51(26):4719–4721.
- Frank PA, Hodgson RH, Comes RD. 1963. Evaluation of herbicides applied to soil for control of aquatic weeds in irrigation canals. *Weeds* 11:124–128.
- Getsinger KD, Madsen JD. 2021. Drawdown herbicide applications for control of flowering rush on dewatered littoral sites. *J. Aquat. Plant Manag.* 59:85–89.
- Getsinger KD, Poovey AG, Glomski L, Slade JG, Richardson RJ. 2011. Utilization of herbicide concentration/exposure time relationships for controlling submersed invasive plants on Lake Gaston, Virginia/North

- Carolina. U.S. Army Corps of Engineers, Engineer Research and Development Center, Environmental Laboratory, Tech. Rep. ERDC/EL TR-11-5. 76 pp.
- Grey TL, Bridges DC, Prostko EP, Eastin EF, Johnson WC, Vencill WK, Brecke BJ, MacDonald GE, Tredaway-Ducar JA, Everest JW, Wehtje GR, Wilcut JW. 2003. Residual weed control with imazapic, diclosulam, and flumioxazin in southeastern peanut (*Arachis hypogaea*). *Peanut Sci.* 30(1):22–27.
- Grichar WJ. 2006. Weed control and grain sorghum tolerance to flumioxazin. *Crop Protect.* 25(2):174–177.
- Grossmann K, Ehrhardt T. 2007. On the mechanism of action and selectivity of the corn herbicide topramezone: A new inhibitor of 4-hydroxyphenylpyruvate dioxygenase. *Pest Manag. Sci.* 63(5):429–439.
- Guardo M, Fink I, Fontaine T, Newman S, Chimney M, Bearzotti M, Goforth G. 1995. Large-scale constructed wetlands for nutrient removal from storm water runoff: An Everglades restoration project. *Environ. Manag.* 19:879–889.
- Haller WT, Sutton DL. 1975. Community structure and competition between hydrilla and vallisneria. *Hyacinth Contr. J.* 13:48–50.
- Harlan SM, Davis GJ, Pesacreta GJ. 1985. Hydrilla in three North Carolina lakes. *J. Aquat. Plant Manag.* 23:68–71.
- Harmanci P, Yaman E, Kaya MD. 2024. A promising method for selecting imazamox-resistant sunflower plants. *Int. J. Agric. Environ. Food Sci.* 8(2):479–485.
- Harrington TB, Minogue PJ, Lauer DK, Ezell AW. 1998. Two-year development of southern pine seedlings and associated vegetation following spray-and-burn site preparation with imazapyr alone or in mixture with other herbicides. *New Forests* 15:89–106.
- Havens KE, Fox D, Gornak S, Hanlon C. 2005. Aquatic vegetation and largemouth bass population responses to water-level variations in Lake Okeechobee, Florida (USA). *Hydrobiologia* 539:225–237.
- Havens KE, Sharfstein B, Brady MA, East TL, Harwell MC, Maki RP, Rodusky AJ. 2004. Recovery of submerged plants from high water stress in a large subtropical lake in Florida, USA. *Aquat. Bot.* 78:67–82.
- Heman ML, Campbell RS, Redmond LC. 1969. Manipulation of fish populations through reservoir drawdown. *Trans. Amer. Fish. Soc.* 2:293–304.
- Herrmann CM, Goll MA, Phillipon CJ, Zandstra BH. 2017. Postemergence weed control in onion with bentazon, flumioxazin, and oxyfluorfen. *Weed Technol.* 31(2):279–290.
- Hestand RS, Carter CC. 1974. The effects of a winter drawdown on aquatic vegetation in a shallow reservoir. *Hyacinth Contr. J.* 12:9–12.
- Hestand RS, Carter CC. 1975. Succession of aquatic vegetation in Lake Ocklawaha two growing seasons following a winter drawdown. *Hyacinth Contr. J.* 13:43–47.
- Hiltibran RC. 1966. Results of pre-emergent aquatic herbicides in Illinois. Abstract. *Weed Society of America* 100-1. <https://www.cabidigitallibrary.org/doi/full/10.5555/19682300273>. Accessed 29 August 2025.
- Hinman EH, Hess AD. 1949. Biological activities of the public health program of the Tennessee Valley Authority. *J. Tenn. Acad. Sci.* 24:195–205.
- Islam MS, Hunt TD, Liu Z, Butler KL, Dugdale TM. 2018. Sediment facilitates microbial degradation of the herbicides endothall monoamine salt and endothall dipotassium salt in an aquatic environment. *Int. J. Environ. Res. Public Health* 15(10):2255.
- Jabusch TW, Tjeerdema RS. 2006. Microbial degradation of penoxsulam in flooded rice field soils. *J. Agric. Food Chem.* 54(16):5962–5967.
- Joshi S, Srivastava A. 2019. Effect of pH on dissipation kinetics of penoxsulam herbicide in water. *Pantnagar J. Res.* 17(3):219–223.
- Kadlec JA. 1962. Effects of a drawdown on a waterfowl impoundment. *Ecology* 43:267–281.
- Kalsi NK, Kaur P. 2019. Dissipation of bispyribac sodium in aridisols: Impact of soil type, moisture and temperature. *Ecotoxicol. Environ. Safety* 170:375–382.
- Kaur H, Kaur P. 2018. Effect of soil type, moisture and temperature on the dissipation of penoxsulam in soil under laboratory conditions. *Bull. Environ. Contam. Toxicol.* 101:803–809.
- Lycan DW, Hart SE. 2006a. Foliar and root absorption and translocation of bispyribac-sodium in cool-season turfgrass. *Weed Technol.* 20(4):1015–1022.
- Lycan DW, Hart SE. 2006b. Cool-season turfgrass reseeding intervals for bispyribac-sodium. *Weed Technol.* 20(2):526–529.
- Madsen JD, Woolf TE, Wersal RM. 2017. Flowering rush control on drawdown sediment: Mesocosm and field evaluations. *J. Aquat. Plant Manag.* 55:42–45.
- Mahoney KJ, Shropshire C, Sikkema PH. 2014. Weed management in conventional- and no-till soybean using flumioxazin/pyroxasulfone. *Weed Technol.* 28(2):298–306.
- Mann RK. 2015. Development and global utility of penoxsulam in rice. pp. 104–105. In: 25th Asian-Pacific Weed Science Society Conference on “Weed Science for Sustainable Agriculture, Environment and Biodiversity,” Hyderabad, India.
- Manning JH, Johnson RE. 1975. Water level fluctuation and herbicide application: An integrated control method for hydrilla in a Louisiana reservoir. *Hyacinth Contr. J.* 13:11–17.
- Manning JH, Sanders DR. 1975. Effects of water level fluctuation on vegetation in Black Lake, Louisiana. *Hyacinth Contr. J.* 13:17–23.
- Mendiburu FD. 2019. *Agricolae*: Statistical procedures for agricultural research. R package version 1.3-1. <https://CRAN.R-project.org/package=agricolae>.
- Miller MR, Norsworthy JK. 2018. Assessment of florypyrauxifen-benzyl potential to carryover to subsequent crops. *Weed Technol.* 32(4):404–409.
- Monika SA, Suyal A, Srivastava PC. 2017. Persistence behavior of penoxsulam herbicide in two different soils. *Bull. Environ. Contam. Toxicol.* 99(4): 470–474.
- Montgomery M, Yu TC, Freed VH. 1972. Kinetics of dichlobenil degradation in soil. *Weed Res.* 12(1):31–36.
- Mueller TC, Boswell BW, Mueller SS, Steckel LE. 2014. Dissipation of fomesafen, saflufenacil, sulfentrazone, and flumioxazin from a Tennessee soil under field conditions. *Weed Sci.* 62(4):664–671.
- Nagid EJ, Tuten T, Johnson KG. 2015. Effects of reservoir drawdowns and the expansion of hydrilla coverage on year-class strength of largemouth bass. *N. Am. J. Fish. Manag.* 35:54–61.
- Nelson LS, Van TK, Lembi CA, Chand T. 1991. Plant growth regulators for aquatic plant management. p. 221. In: Proceedings of the 25th Annual Meeting, Aquatic Plant Control Research Program, Orlando, FL. U.S. Army Corps of Engineers, Waterways Experiment Station. Miscellaneous Paper A-91-3.
- Netherland MD. 1997. Turion ecology of hydrilla. *J. Aquat. Plant Manag.* 35:11–10.
- Nichols SA. 1975. The use of overwinter drawdown for aquatic vegetation management. *Water Resour. Bull.* 11:1137–1148.
- Pannacci E, Onofri A, Covarelli G. 2006. Biological activity, availability and duration of phytotoxicity for imazamox in four different soils of central Italy. *Weed Res.* 46(3):243–250.
- Patten K. 2003. Persistence and non-target impact of imazapyr associated with smooth cordgrass control in an estuary. *J. Aquat. Plant Manag.* 41:1–6.
- Poovey AG, Kay SH. 1998. The potential of a summer drawdown to manage monoeocious hydrilla. *J. Aquat. Plant Manag.* 36:127–129.
- R Core Team. 2019. R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna, Austria. <https://www.R-project.org/>.
- Rahman A, Dowsett C, Trolove M, Trevor J. 2015. Soil residual activity and plant-back periods for the herbicides saflufenacil and topramezone. *Bioeconomy Science Institute, AgResearch Group.* <https://hdl.handle.net/11440/1228>.
- Ramanathan SS, Gannon TW, Locke AM, Everman WJ. 2023. Characterizing atrazine, mesosulfuron-methyl, and topramezone bioavailability in North Carolina soils using greenhouse bioassays. *Agrosyst. Geosci. Environ.* e20371.
- Reimer DN. 1984. Introduction to freshwater vegetation. AVI Publishing Co. Westport, CT, USA. 207 pp.
- Richardson LV. 1975. Water level manipulation: A tool for aquatic weed control. *Hyacinth Contr. J.* 13:8–11.
- Richardson RJ, Zandstra BH. 2006. Evaluation of flumioxazin and other herbicides for weed control in gladiolus. *Weed Technol.* 20(2):394–398.
- Richardson RJ, Zandstra BH. 2009. Weed control in Christmas trees with flumioxazin and other residual herbicides applied along or in tank mixtures. *HortTechnol.* 19(1):181–186.
- Rood J, Campbell J, Thill D, Ball D, Bennett L, Yenish J, Nelson J, Rood R, Shafii B, Price W. 2012. Tillage affects imazamox carryover in yellow mustard. *Weed Technol.* 26(2):308–315.
- Sartain BT. 2014. Developing management recommendation for hydrilla (*Hydrilla verticillata* L.f. Royle) in the Ross Barnett Reservoir: A community approach. Master's thesis. Mississippi State University, Starkville, MS. 117 pp.

- Schreiber F, Scherner A, Massey JH, Zanella R, Avila LA. 2017. Dissipation of clomazone, imazapyr, and imazapic herbicides in paddy water under two rice flood management regimes. *Weed Technol.* 31(2):330–340.
- Sharip Z, Schooler SS, Hipsey MR, Hobbs RJ. 2012. Eutrophication, agriculture and water level control shift aquatic plant communities from floating-leaved to submerged macrophytes in Lake Chini, Malaysia. *Biol. Invas.* 14:1029–1044.
- Shearer JF, Nelson LS. 2009. Combining ALS-inhibiting herbicides with the fungal pathogen *Mycropleptodiscus terrestris* for control of hydrilla. U.S. Army Corps of Engineers, Engineer Research & Development Center. Tech. Note APCRP-CC-11. 9 pp.
- Simsman GV, Chester G. 1975. Persistence of endothall in the aquatic environment. *Water Air Soil Pollut.* 4:399–413.
- Steward KK. 1969. Effects of growth regulators and herbicides on germination of hydrilla turions. *Weed Sci.* 17(3):299–301.
- Steward KK. 1980. Retardation of hydrilla (*Hydrilla verticillata*) regrowth through chemical control of vegetative reproduction. *Weed Sci.* 28(3):245–251.
- Sutton DL, Portier KM. 1985. Density of subterranean turions and turions of hydrilla in South Florida. *J. Aquat. Plant Manag.* 23:64–67.
- Taylor JL. 1966. CASORON, a new aquatic herbicide. *Hyacinth Contr. J.* 5:20–21.
- Taylor JL. 1967. Control of chara with CASORON AQ. *Hyacinth Contr. J.* 6:19.
- Teuton TC, Main CL, Sorochan JC, McElroy JS, Mueller TC. 2007. Annual bluegrass (*Poa annua*) control in creeping bentgrass (*Agrostis stolonifera*) putting greens with bispyribac-sodium. *Weed Technol.* 21(2):426–430.
- Turgeon AJ, Meggitt WF, Penner D. 1972a. Role of endothall in the control of annual bluegrass in turf. *Weed Sci.* 20(6):562–565.
- Turgeon AJ, Penner D, Meggitt WF. 1972b. Selectivity of endothall in turf. *Weed Sci.* 20(6):557–561.
- Turra GM, Cutti L, Machado FM, Dias GM, Andres A, Markus C, Merotto A. 2023. Application of ALS inhibitors at pre-emergence is effective in controlling resistant barnyardgrass biotypes depending on the mechanism of resistance. *Crop Prot.* 172:106325.
- Ulbrich AV, Souza JRP, Shaner D. 2005. Persistence and carryover effect of imazapic and imazapyr in Brazilian cropping systems. *Weed Technol.* 19(4):986–991.
- Van T, Steward K. 1990. Longevity of monoecious hydrilla propagules. *J. Aquat. Plant Manag.* 28:74–76.
- Van Geest GJ, Coops H, Roijackers RMM, Buijse AD, Scheffer M. 2005. Succession of aquatic vegetation driven by reduced water-level fluctuations in floodplain lakes. *J. Appl. Ecol.* 42:251–260.
- Vassios JD. 2010. Evaluation of herbicides for control of Eurasian watermilfoil and sago pondweed. Master's thesis, Colorado State University, Fort Collins, CO. 51 pp.
- Wersal RM, Madsen JD, Gerard PD. 2013. Survival of parrotfeather following simulated drawdown events. *J. Aquat. Plant Manag.* 51:22–26.
- Wood JD, Netherland MD. 2019. Laboratory response of monoecious *Hydrilla* to four slow-acting, enzyme-inhibiting herbicides. *J. Aquat. Plant Manag.* 57:38–41.
- Wright HE, Norsworthy JK, Roberts TL, Scott RC, Hardke JT, Gbur EE. 2020. Use of floraspyrauxifen-benzyl in non-flooded rice production systems. *Crop Forage Turfgrass Manag.* 7(1):e20081.
- Zaccaro MLM, Burd JD, Russell DP. 2018. Tolerance of several legumes to residual imazapyr applied under greenhouse conditions. *Weed Technol.* 32(1):66–71.